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Cold Stress in Rice at Early Growth Stage – An Overview

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ABSTRACT

The changing climate accompanied with cold temperature which unmasks itself as a devastating stress for most of the high yielding rice cultivars in twenty first century. The early growth stage which include germination, seedling formation get severely damaged by cold stress leading into crop failure or major yield loss. However, the genotypes grown in colder climates of high altitude and temperate regions have evolved themselves to avoid or survive cold stress in course of time. In order to achieve cold tolerance in high yielding rice, a thorough understanding of this tolerance mechanism at physiological, metabolic and genomic level is imperative. So, the present review on rice covers the cold damage evaluation, characterization, physiological response to stress and tolerance mechanism in brief.

Key words: Rice, Early stage, Seedling, Cold, Stress, Tolerance

INTRODUCTION

Rice (Oryza sativa L.) is an annual grass which belongs to the family Gramineae, subfamily Poaceae². It is one of the earliest domesticated and principal staple food crop for more than one third of the worlds' population. In Asia, more than 90% of the worlds' rice is grown, where almost 60% of the worlds' population lives⁴. Abiotic stress related with temperature due changing to climatic conditions of twenty first century is the burning concern for plant scientists worldwide^{68,82}. Low temperature in particular has alarming consequences on plant growth and development, since these processes have optimum temperature requirements specific for each and every plant species.

Low temperatures pose a major climatic problem for all rice growing countries including Australia, China, Japan, Nepal, Russia and South Korea¹⁷. The Australian rice industry, which has the highest yield (10 t h⁻¹) in the world experiences cold stress once in about every 4 years and resulted in about 30 - 40 % yield loss accounting \$120 million due to cold-induced sterility⁵⁵.

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In India, cold rice is grown in about 1 million ha of hill regions in Jammu and Kashmir, Uttaranchal and North eastern hill states accounting 2.3% of total area under rice. The average yield of this cold rice is about 1.1 t h⁻¹ as against the average national yield of 9.1 t h ¹. Major productivity constraints of these areas include low temperature, blast, drought spell and very short span of cropping season. The cold temperature stress in those areas affect rice mostly at early stage and seldom at the flowering stages too, resulting in sterility and devastating yield reduction. Since, rice is a temperature-sensitive field crop, temperature induced yield loss is a worldwide problem⁶³.

Types of Low Temperature Injury

A plant can endure through two types of injuries after low temperature exposure⁷⁸. 1) due to lower Chilling injury occurs temperature just above freezing point of water. This phenomenon remains reversible initially but ultimately causes cell death due to prolonged cold spell. Sometimes progressive colder temperature beyond critical range may result in hardening and/or acclimatization of plants which can lessen and/or abolish stress injury. 2) Freezing injury is induced by the low temperature below freezing point. The intracellular freezing becomes fatal for the protoplasmic structure when the ice crystals grow large enough to disrupt the cells. In extra cellular freezing, the protoplasm of the plant becomes dehydrated because a water vapour deficit is created as cellular water is transferred to ice crystals formed in the intercellular spaces. Rice crop is

more commonly abused by the first kind of injury, i.e. chilling injury, whereas, freezing injury may also occur once in a while⁷⁸.

Tools To Evaluate Cold Stress in Rice

Most of the physiological assessment of rice in terms of cold tolerance or sensibility have been made in two phases of growth: seedling and booting. In both of them cold temperature has harmful effects on crop productivity, as in the first one the number of established plants is affected and in the booting stage pollen sterility is induced by cold, decreasing the final number of grains. A wide range of practices such as altered cold intensities and episodes of exposure are usually applied to assess the cold injury and tolerance in these developmental phases. Only a few of them are non-destructive. Primarily, the visual symptoms as wilting and yellowing of leaves at seedling stage are correlated with cold stress in general^{73,77,89}. So, the degree of leaf withering is used as an essential measure for recording chilling damage⁵⁴. Chilling injury and low temperature chlorosis is used as a scoring measure for cold tolerance/susceptibility during seedling stage.

Moreover, seedling survival is also used as a scoring measure for cold tolerance, since susceptible seedlings struggle in upholding average metabolic rates under cold and eventually die⁵². There are standard screening methodologies (Table 1) described by several scientists to screen rice genotypes effectively for cold response under controlled temperature conditions at germination (seedling) stage are given below.

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Table 1: Methods and traits evaluated in germination stage for cold tolerance selection in rice

	<u> </u>		
Methodology of screening	Evaluated trait	Reference	
10, 15, 20, and 25°C for 5 to 30 days (on the	Germination rate (radicle	Bertin et al ⁷	
basis of temperature)	protrusion)		
13 °C for 28 days and 28 °C for 7 days	Germination index and	Priyanka <i>et al</i> ⁶⁴	
13 C for 28 days and 28 C for 7 days	scoring for cold tolerance 1-9		
17°C for 7 days	Germination (%) and its speed	Sthapit and Witcombe ⁷⁶	
13°C to 15°C for 7 days	Percentage of germination		
150C for 12 16 done	Low temperature	Sheng et al ⁷⁰	
15°C for 12-16 days	germinability (LTG)		
15°C for 10 days	Coleoptile length	Hou et al ²⁵	
15°C for 6 days	Germination rate	Chen et al ¹⁰	
13/20°C day/night and control condition	Leaf discoloration, SPAD	Park et al ⁶²	
25/20°C day/night15 days	value chlorophyll content		
12-h light (15000 LX) 12-h dark. The			
seedlings were initially exposed to 14°C for 2	Seedling sensitive	Yang et al ⁹⁰	
h followed by 12°C for 4 h and 10°C for 4 h.			
10 °C for 10 days, 10 °C for 13 days	Seedling survival percentage	Zhang et al ⁹⁶	
12°C. for 10 days	Seedling vigor	Han et al ²²	
4°C for 48 h	Cold induced injury	Xiao et al ⁸⁴	
25 °C for 4 days	Dormancy	Xie et al ⁸⁶	
18–19°C cold-water irrigation (field) 17–	G 11	133	
18°C cold-air (glasshouse)	Cold sensitive	Jena et al ³³	
4°C for 9 days, 4°C for 11 days and 4°C	Cold stress tolerance index	Juan et al ³⁵	
for14 days	and withering index		
1400 0 7 1 11 1 14 1 117 1	Low temperature vigor of	Han et al ²³	
14°C for 7 d, 11 d, 14 d, and 17 d	germination (LVG)		
50°C for 48 h to break dormancy, 32°C for			
36 h, 5°C for 10 days, 20°C for 10 days to	Seedling survival	Pan et al ⁶¹	
recover			
12°C for 28 days 28°C for 7 days	Coleoptile and radicle length	Bosetti <i>et al</i> ⁹	
13°C for 28 days, 28°C for 7 days	under Bosetti et at		
13°C for 28 days, 28°C for 7 days	Radicle and coleoptile length	Dashtmian <i>et al</i> ¹⁵	
15 C 101 20 days, 20 C 101 / days	and germination index	Dashtillali et al	

Most of the reported studies have mentioned the seedling survival rate of transgenic plants as baseline criteria for tolerance to cold stress 11,26,27,28,31,39,46,47,50,73,77,87,88,89,95. However, an on/off character like survival proved to be incompetent in the actual field condition since cold tolerance is a variable trait, and it is not even a useful trait for QTL detection. In few of the other studies, quantitative analysis of shoot and root biomass were also used for evaluation of contrasting genotypes and/ or transgenic plants 19 at any developmental stages, since plant growth and development is often negatively subjected to cold stress 66. However,

most of the evaluation methods have the drawback of being destructive, time intense and are not suitable for plant breeding curriculums where huge number of lines in the field need to be assessed.

Low Temperature at Germination Stages

Seed germination is the most important stage in plant life cycle where thrusts the seed to grow into a plant run and produces many seeds. Conditions essential for germination are water, air, temperature and light. In case of rice, quiescent as well as non-dormant seeds just need rehydration after proclamation from primary dormancy. However, the germination

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of rice seed is affected greatly by temperature. The temperature colder than the favourable range⁵⁷ (18°C to 33°C) retard the germination progression starting from imbibition, activation and succeeding manifestation.

Cold temperature slows down the diffusion resulting in disrupted imbibition process and escape of solutes from the seeds. However, Yoshida⁹³ reported the successive stages of germination (i.e. growth of coleoptile and radical) as the most vulnerable phases to cold spell. It (cold stress) results in retarded

cell division and cell elongation in plants because of unbalanced metabolic activities at such low temperature⁴⁹. Rice is a tropical and/or sub-tropical plant which requires a fairly high temperature ranging from 20°C to 40°C⁷⁵. The standard temperature for rice seed germination is considered to be approximately 30°C. The temperature below 20°C results in gradual decrease of germination rate⁸⁰ (**Table 5**). Moreover, Yoshida⁹⁴ considered 10°C as the minimum critical temperature of rice germination.

Table 5: Temperature and seed germination

	30°C	→ Optimum	→ Very good germination
emp	20°C	> Critical	Medium Germination
I	10°C	→ Limiting	Germination Failure

Cold stress in rice delays germination and emergence; soil temperature of below 10°C can result in complete failure of germination⁹⁴. Screening for cold tolerance based on germination and seeding growth have been attempted in rice as well¹³ and there was marked genetic variability for the traits⁶⁷. Yoshida⁹⁴ studied the effect of cold stress at phases; germination, imbibition. activation and post germination growth. The effect of cold stress was more pronounced at the phase of imbibing and this was regarded as the most sensitive phase. The exposure of seeds to cold stress during this phase has resulted in increased escape of solutes from the seeds. This has been attributed to the incomplete plasma membrane of the dry seed the disturbance caused reconstruction¹². Cold stress at this stage has been reported to target the cellular membrane and thus is the primary cause of other metabolic disorders usually observed within the cells⁴⁹. Despite the fact that seed germination under low temperature is a main

problem in rice plant, there's still an extended manner to go to elucidate the mechanism of seed germination. Lately, purposeful genomic techniques have been implemented to observe and to elucidate the mechanism of seed germination in rice.

Low Temperature at Seedling Stage

Plants require an optimal temperature range for their growth, development and ultimate survival⁴⁵. Low temperature has a strong impact on growth, survival, reproduction and distribution of plants. The seedlings get severely damaged by cold stress when they are grown in winter environments. As a result, the productivity decreases in temperate areas 50,85. In the Northern and North-eastern parts of India, cold spell prevails in winter season during December to February and the minimum temperature remains often below 12-15°C. Infact, the minimum temperature occasionally reaches below 20°C during March and April in some parts of the northern states of India.

Even though, rice is being cultivated in a wide range of environments (tropical, sub-tropical, temperate), it is still a tropical C₃ crop³⁸, which yields best under warm temperatures and high solar radiation¹⁶. It is been reported^{53,65} to be more sensitive to cold stress than any other cereal crops, especially during seedling, tillering, panicle development and flowering stages. The critical temperature for rice growth varies with different developmental phases such as 10°C for germination and 17°C for the reproductive stages. Temperature drops to about 10°C during seedling establishment (October to early November) such low temperature significantly reduces seedling growth and establishment³⁰. Nishiyama⁵⁹ and Yoshida⁹² also reported that the critical low temperature differs according to variety, duration of low temperature and the plant's physiological development. Nakagahra et al⁵⁶.. Sharifi⁶⁹ and Lou et al^{48} , found that rice is vulnerable to damage by temperatures below 15°C, especially, the early seedling phase which is the foundation for stable seedling formation following strong vegetative growth. Induction of colder temperature during seedling stage results in lower number of seedlings, shrinked tillering⁷¹, higher plant mortality^{6,19,51}, and also induce non-uniform crop maturity⁷².

The seedling growth drop in rice due to low temperature poses a major threat in tropical and sub-tropical zones at high elevation as well as regions where cold mountain water is used for irrigation. In such areas, water temperature below 15° C causes germination, delay in seedling emergence, poor seedling establishment, slow growth, yellowing and drying of leaves, reduced tillering which ultimately lead to seedling death^{36,51,56}. Cold stress also affects chlorophyll content and thus interferes in photolysis^{37,40}. Most of the high yielding varieties cannot be used in direct sowing because of low germination rate at low temperature.

Bardhan and Biswas⁵ reported the adversely affects of low temperature on seedling dry matter since low temperature affects the photosynthetic activity. Dai et at^{14} ., reported a positive relationship between root oxidizing activity and dry matter production in rice seedlings under cold stress which also could be used as a criteria for cold tolerant rice variety selection.

Table 1: Response of the rice plant to varying temperature at different stages

Growth stage	Critical temperature		
	Low	Optimum	High
Germination	16-19	18-40	45
Seedling emergence and establishment	12-35	15-30	35
Rooting	16	25-28	35
Leaf elongation	7-12	31	45
Tillering	9-16	25-31	33
Initiation of panicle primordia	15	-	-
Panicle differentiation	15-20	-	30
Anthesis	22	30-33	35-36
Ripening	12-18	20-29	> 30

Physiological Response of Rice to Low Temperature

Low temperature adversely affects a wide range of physiological processes from seed germination to maturity, and ultimately causes a serious yield reduction in rice. Changes in physiological activities precede development of visual symptoms. At colder temperature, the rate of respiration and ion leakage increase; but the photosynthetic activity and carbohydrate metabolism decrease. Nutrient deficiency at root is one of the consequences of low temperature²⁰. Low temperature induces chlorosis in developing young leaves, which remain white, even under permissive temperature conditions, without withering⁹¹. Chilling temperature even induce starch build-up in chloroplast which hinders photosynthetic activities due to feedback inhibition of photosynthetic enzymes. Photoinhibition of photosynthetic machinery is enhanced at chilling temperatures due to decreased utilisation of excitation energy⁷⁴. Several factors such as type of plant species, developmental stage, nutrition, irradiance and other climatic conditions before, during and after the chilling exposure influences the degree of dysfunction. Rice genotypes belonging to indica subspecies are known to be more sensitive to low temperatures than iaponica rice^{12,24}. Early germination and microspore genesis stages in rice are considered as the most sensitive to low temperature stress^{8,12,21}. Low temperature at seedling and / or panicle development stage is greatly exacerbated by increased nitrogen application. Chilling causes greater injury under illuminated conditions, owing to photo oxidation damage determined by the increase of harmful active oxygen species^{41,83}. Cloudy conditions or darkness during chilling stress reduced the damage in photosynthetic machinery.

Mechanism of Cold Tolerance in Rice

Low temperature or cold tolerance is the ability of plants to survive and perform under cold stress conditions. The capacity to maintain total plant yield is a tolerance criteria considered by several workers. It allows all possible yield compensatory mechanisms to occur during the vegetative and reproductive period. Rice plants have evolved various survival mechanisms (tolerance) through changes in their morphophysiological and biochemical behaviour.

It has been found that sucrose and other simple sugars accumulate during the acquisition of cold tolerance in most species and is believed to be instrumental in protecting membranes during freezing, but this alone is not sufficient to confer full tolerance. This fact is presage by the bewildering complexity of changes in gene expression that accompanies the acquisition of cold and freezing tolerance in species capable of cold acclimation⁷⁸. Although the actual function of most of these cold-induced genes in the development of cold tolerance is unknown. It is known that many cold-regulated genes contain a drought responsive DNA element (ORE) that interacts with the transcriptional activator CBF1 (C-Binding Factors)⁷⁸. Repeat Constitutive expression of CBF1 confers considerable cold/freezing tolerance to unacclimatized plants confirming its central role in the acclimatization process^{32,34} although it is also not alone sufficient for full development of cold acclimatization. Thus, while native respond to temperate plants repeated cool temperature exposures to progressively acquiring greater tolerance through intricate if only partially understood acclimation process, warm climate species respond by accumulating damage progressively becoming physiologically unfit.

The modest level of genetic variation for chilling tolerance that does exist in these warm climate species appears to reside in their recovery ability rather than in their acclamatory capacity.

The morphological adaptations with decreasing temperature are often linked to decrease in leaf area ratio (LAR), specific leaf area (SLA) and relative growth rate (RGR). Increase in leaf thickness is an adaptation mechanism to protect the photosynthetic machinery against the cold⁸¹. Rice genotypes having high respiratory homeostasis (H), *i.e.* an ability of the plants to maintain similar respiration rates at growth temperatures, showed greater tolerance and maintained both shoot and root growth under cold conditions.

Plants have also developed antioxidant systems which shield cellular organelles and membranes from abating effects of cold oxygen species^{18,42}. induced reactive Antioxidant enzymes, such as SOD, CAT and POX, can react with, and neutralize the activity of AOS⁶⁰. In conjunction with these enzymes antioxidant compounds such as ascorbate, glutathione, etc. also help in removal of noxious oxygen compounds⁸³. Cold tolerant rice cultivars have been reported to maintain higher activities of defence enzymes and higher content of antioxidants with little effect on electrolyte leakage^{29,42}.

There are certain organic compounds called osmoprotectants which accumulate in plants during acclimatization in continuously altering environment. These include amino acid (proline), betaine (), sugar (trehalose) and polyamines such as spermine, spermidine and putrescine. In addition to the direct protective effects, osmoprotectants are also involved in the up regulation of several genes (new proteins are synthesized) such as *Weor* 410 and *Weor* 413³. Except glycine betaine, all

other osmoprotectants occur in rice tissues at low levels. Cold tolerant cultivars, which accumulate higher spermine, produce more viable pollen grains leading to low spikelet sterility⁵⁸. Higher gibberellic acid (GA) content in the seedlings resulted in greater seedling cold tolerance (biomass production) in rice cultivars. However, GA content in the leaves at reproductive stage could not be significantly correlated with spikelet sterility⁵⁵. studies on production accumulation of organic compounds (osmoprotectants) in rice cultivars under cold temperatures lead⁵⁵ to present the following mechanism for low temperature tolerance in rice.

Higher GA production leads to greater α-amylase activity thus liberating simple sugars for the growth of the germinating seeds. On the other hand, osmoprotectants such as spermine could protect activity of enzymes such as α -amylase, various physiological processes including photosynthesis, and could enhance the development of engorged prolin A. A similar association has been shown between high proline content in pollen and fertility⁴³ pollen have high recently demonstrated the importance of spermine in drought tolerance in rice. Rice plants engineered to produce higher spermine were able to tolerate drought through antisenescence compared to the wild type plants with lower capacity to accumulate spermine.

Future Directions

So far, the scientists have only approached to identify individual types of responses in rice due to cold stress. Different types of alterations in those responses by using breeding and genome modification techniques have already provided with expected level of tolerance. However, those approaches have also resulted in unwanted collateral translates

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or missing translational products along with the expected one. In the bigger picture of cold tolerance and susceptibility, a sustainable outcome is possible only if the researchers consider all the regulatory mechanisms and pathway in a single event of approach, when only the targeted phases are being affected without influencing the rest of the off target phases. The revolution in modern molecular biological techniques such as the genome editing with CRISPR/cas system, meganucleases sound promising in such endeavours of twenty first century.

REFERENCES

- 1. Aghaee, A., Moradi, F., Zare-Maivan, H., Zarinkamar, F., Pour Irandoost, H. and Sharifi, P., Physiological responses of two rice (Oryza sativa L.) genotypes to chilling stress at seedling stage. Afr. J. Biotechnol., **10:** 7617–7621 (2011).
- 2. Agropedia ., Botanical classification of Rice, Submitted by naipictuasd harwad on 06/03/2009 Friday. -12:51. http://agropedia.iitk.ac.in/content/botanical classification-rice (2009).
- 3. Allard, F., Houde, M., Krol, M., Ivanov, A., Huner, N.P.A. and Sarhan, F., Betaine improves freezing tolerance in wheat. Plant Cell Physiology, 39: 1194-1202 (1998).
- 4. Ashkani Sadegh, Mohd Y. Rafii , Mahmoodreza Shabanimo frad, Gous Miah, Mahbod Sahebi, Parisa Azizi, Fatah A.Tanweer, Mohd Sayeed Akhtar and Abbas Nasehi, Molecular breeding strategy and challenges towards improvement of blast disease resistance in rice crop. Frontiers in Plant Science, 6: 886 (2015).
- 5. Bardhan Roy, S.K. and Biswas, S., Low temperature effect on rice seedlings. Oryza 20: 204-208 (1983).
- 6. Baruah, A.R., Ishigo-Oka, N., Adachi, M., Oguma, Y., Tokizono, Y., Cold tolerance at the early growth stage in wild and

- cultivated rice. Euphytica, 165: 459-470 (2009).
- 7. Bertin, P., Kinet, J.M., Bouharmont, J., Evaluation of chilling sensitivity in varieties. different rice Relationship between screening procedures applied during germination and vegetative growth. Euphytica, 89: 201-210 (1996).
- 8. Blum, A., Cold resistance. In Plant Breeding for stress environments. Boca Raton: CRC, cap. 5 pp 99-132 (1988).
- 9. Bosetti, F., Montebelli, C., Dionísia, A., Novembre, L.C., Chamma, H. Pinheiro, J.B., Genetic variation germination cold tolerance in Japanese rice germplasm. Breeding Science, 62: 209-215 (2012).
- 10. Chen, L., Lou, Q., Sun, Z., Xing, Y., Yu, X. and Luo, L., QTL mapping of low temperature on germination rate of rice. Rice Science, 13: 93-98 (2006).
- 11. Chen, N.A., Y. Xu, X. Wang, C. Du, J., Du, M., Yuan, et al., OsRAN2, essential for mitosis, enhances cold tolerance in rice by promoting export of intranuclear tubulin and maintaining cell division under cold stress. Plant Cell Environ., 34: 52-64 (2011).
- 12. Cruz, R.P. and Milach, S.C.K., Cold temperature tolerance at the germination stage of rice: Methods of evaluation and characterization of genotypes. Sci. Agric., **61:** 1–8 (2004).
- 13. Cruz, R.P., Milach, S.C.K. and Federizzi, L.C., Rice cold tolerance at reproductive stage in a controlled environment. Sci. *Agric.*, **63:** 255–261 (2006).
- 14. Dai, L., Lin, X., Ye, C., Ise, K., Saito, K., Kato, A., Xu, F., Yu, T. and Zhang, D., Identification of quantitative trait loci cold tolerance controlling at the reproductive stage in Yunnan landrace of rice. Kunmingxiaobaigu. Breeding Science, 54: 253-258 (2004).
- 15. Dashtmian, F.P., Hosseini, M.K. and Esfahani, M., Methods for rice genotypes cold tolerance evaluation at germination stage. International Journal of Agriculture and Crop Sciences, 5: 2111-2116 (2013).

- 16. Ehleringer, J.R. and Monson, R.K., Evolutionary and ecological aspects of photosynthetic pathway variation. *Annu Rev Ecol Syst*, **24:** 411–439 (1993)
- 17. Farrell, TC., Fox, K.M., Williams, R.L. and Fukai, S., New screening method for cold tolerance during the reproductive stage in rice. http://www.cropscience.org.au/icsc2004/poster/217/1/695-farrellt.htm. (2004).
- 18. Foyer, C.H., Lelandais, M., Galap, C. and Kunert, K.J., Effect of elevated cytoso!icglutathione reductase activity on the cellular glutathione pool and photosynthesis in leaves under normal and stress conditions. *Plant Physiology*, **97**: 863-872 (1991).
- 19. Fujino, K., Sekiguchi, H., Sato, T., Kiuchi, H., Nonoue, Y., Takeuchi, Y., Ando, T., Lin, S.Y. and Yano, M., Mapping of quantitative trait loci controlling low-temperature germinability in rice (Oryza sativa L.). *Theoretical and Applied Genetics*, **108**: 794-799 (2004).
- 20. Gagoi, N. and Baruah, K.K., Effect of cold hardening and GA3 on growth and yield of bora rice. *Indian Journal Plant Physiology*, **5(NS):** 339-343 (2000).
- 21. Gunawardena, T.A, Fukai, 5., arid Blamey, F.P.C., Low temperature-induced spikelet sterility in rice. II. Effect of panicle and root temperatures. *Australlian Journal of Agricultural Research*, **54:** 947-956 (2003).
- 22. Han, L.Z., Qiao, Y.L., Zhang, S.Y., Zhang, Y.Y., Cao, G.L., Kim, J.W., Lee. K. and Koh, H.J., Identification of quantitative trait loci for cold response of seedling vigor traits in rice. *Journal of Genetics and Genomics*, 34: 239-246 (2007).
- 23. Han, L.Z., Zhang, Y.Y., Qiao, Y.L., Cao, G.L., Zhang, S.Y., Kim, J.H. and Koh, H.J., Genetic and QTL analysis for low temperature vigor of germination in rice. *Acta Genetica Sinica*, 33: 998-1006 (2006).
- 24. Hetherington, S.E., He, J. and Smilie, R.M., Photoinhibition at low temperature

- in chilling –sensitive and -resistant plants.

 Plant Physiology, **90:** 1609-1615 (1989).
- 25. Hou, M.Y., Jiang, L., Wang, C.M., Wan, J.M., Detection and analysis of QTL for low temperature germinability in rice (Oryza sativa L.). *Rice Genetics Newsletter*, **20:** 52-55 (2003).
- 26. Hu, H., You, J., Fang, Y., Zhu, X., Qi, Z. and Xiong, L., Characterization of transcription factor gene SNAC2 conferring cold and salt tolerance in rice. *Plant Mol. Biol.* **67:** 169–181 (2008).
- 27. Huang, J., Sun, S.J., Xu, D.Q., Yang, X., Bao, Y.M., Wang, Z.F. et al., Increased tolerance of rice to cold, drought and oxidative stresses mediated by the overexpression of a gene that encodes the zinc finger protein ZFP245. Biochem. Biophys. Res. Commun., 389: 556–561 (2009).
- 28. Huang, J., Sun, S., Xu, D., Lan, H., Sun, H., Wang, Z., *et al.*, A TFIIIA-type zinc finger protein confers multiple abiotic stress tolerances in transgenic rice (Oryza sativa L.). *Plant Mol. Biol.*, **80**: 337–350 (2012).
- 29. Huang, M. and Guo, Z., Response of antioxidative system to chilling stress in two rice cultivars differing in sensitivity. *Biol. Plant.*, **49:** 81-84 (2005).
- 30. Humphreys, L., Sides, R. and Fattore, A., Rice establishment. Farmers' News letter Large Area, **147**: 30-31 (1996).
- 31. Ito, Y., Katsura, K., Maruyama, K., Taji, T., Kobayashi, M., Seki, M. *et al.*, Functional analysis of rice DREB1/CBF-type transcription factors involved in coldresponsive gene expression in transgenic rice. *Plant Cell Physiol.*, **47:** 141–153 (2006).
- Jaglo-Ottosen, K.R., Gilmour, S.J., Zarka, D.G., Schabengerger, O. and Thomashow, M.F., Arabidopsis CBFl over expressioninduces COR genes and enhances freezing tolerance. *Science*, 280: 104-106 (1998).
- 33. Jena, K.K., Kim, S.M., Suh, J.P. and Kim, Y.G., Development of cold-tolerant breeding lines using QTL analysis in rice.

- genetic In: Rice diversity and improvement, 2nd Africa Rice Congress, Bamako, Mali., March 22-26 (2010).
- 34. Jones, L. and Ort, D.R., Circadian regulation of sucrose phosphate synthase aCtivity in tomato by protein phosphatase activity. Plant Physiology, 113: 1167-1175 (1997).
- 35. Juan, J.Z., Xiang, Z.Y., Li, Z.D., Yong, M.L., Ming, L.X., Xin, L.B. and Deng, Y.C., Identification of QTL for rice cold tolerance at plumule and 3-leaf seedling stages by using QTL network software. Rice Science, 17: 282-287 (2010).
- 36. Kaneda, C. and Beachell, H.M., Response of indica-japonica rice hybrids to low temperatures. SABRAO Journal, 6: 17-32 (1974).
- 37. Kanneganti, V. Gupta, and Overexpression of OsiSAP8, a member of stress associated protein (SAP) gene family of rice confers tolerance to salt, drought and cold stress in transgenic tobacco and rice. Plant Molecular Biology, **66:** 445-462 (2008).
- 38. Karki, S., Rizal, G. and Quick, W.P., Improvement of photosynthesis in rice (Oryza sativa L.) by inserting the C4 pathway. Rice 6: 28. doi: 10.1186/1939-8433-6-28. pmid:24280149 (2013)
- 39. Kawakami, A., Sato, Y. and Yoshida, M., Genetic engineering of rice capable of synthesizing fructans and enhancing chilling tolerance. J. Exp. Bot., 59: 793-802 (2008).
- 40. Kim, S.J., Lee, S.C., Hong, S.K., An, K., An, G. And Kim, S.R., Ectopic expression of a cold responsive OsAsr1 cDNA gives enhanced cold tolerance in transgenic rice plants. Molecules and Cells, 27: 449-458 (2009).
- 41. Krishnasamy, V. and Seshu, D.V., Seed germination rate and associated characters in rice. Crop Science, 29: 904-908 (1989).
- 42. Kuk, Y.I., Shin, J.S., Burgos, N.R., Hwang, T.E., Han, O., Cho, B.H., Jungs, S. and Guh, J.O., Antioxidative enzymes offer protection from chilling damage in

- rice plants. Crop Science, 43: 2109-2117 (2003).
- 43. Lansac, A.R., Sullivan, C.V. and Johnson. B.E., Accumulation of free proline in sorghum (Sorghum bicolour) pollen. Canadian Journal of Botany, 74: 40-45 (1996).
- 44. Lee, M.H., Fukai, S. and Basnayake, J., Increased lowland rice production in the Mekong Region. In: Proceedings of an international workshop, Vientiane, Laos, 30 October to 2 November 2000 at Center for International Australian Agricultural Research, Canberra, Australia on Low temperature tolerance in rice: the Korean experience, 109–117 (2001).
- 45. Levitt, J., Responses of Plants Environmental Stresses, 2nd ed. New York, Academic Press. (1980).
- 46. Li, H.W., Zang, B.S., Deng, X.W. and Wang, X.P., Overexpression of the trehalose-6-phosphate synthase gene OsTPS1 enhances abiotic stress tolerance in rice. Planta, 234: 1007-1018 (2011).
- 47. Liu, K., Wang, L., Xu, Y., Chen, N., Ma, Q., Li, F. et al., Overexpression of OsCOIN, a putative cold inducible zinc finger protein, increased tolerance to chilling, salt and drought, and enhanced proline level in rice. Planta, 226: 1007-1016 (2007).
- 48. Lou, Q., Chen, L., Sun, Z., Xing, Y., Li, J., Xu, X., Mei, H. and Luo, L., A major QTL associated with cold tolerance at seedling stage in rice (Oryza sativa L.). Euphytica, **158:** 87-94 (2007).
- 49. Lyons, J.M. Chilling injury in plants. Ann. Rev. Plant Physiol., 24: 445–466 (1973).
- 50. Ma, Q., Dai, X., Xu, Y., Guo, J., Liu, Y. and Chen, N. et al., Enhanced tolerance to chilling stress in OsMYB3R-2transgenic rice is mediated by alteration in cell cycle and ectopic expression of stress genes. Plant Physiol., 150: 244-256 (2009).
- 51. Mackill, D.J. and Lei, X.M., Genetic variation for traits related to temperate adaptation of rice cultivars. Crop Science, **37:** 1340-1346 (1997).

- 52. Morsy, M.R., Jouve, L., Hausman, J.F., Hoffmann, L. and Stewart, J.D., Alteration of oxidative and carbohydrate metabolism under abiotic stress in two rice (Oryza sativa L.) genotypes contrasting chilling tolerance. *J. Plant Physiol.*, **164:** 157–167 (2007).
- 53. Mukhopadhyay, A., Vij, S. and Tyagi, A.K., Overexpression of a zinc-finger protein gene from rice confers tolerance to cold, dehydration, and salt stress in transgenic tobacco. *Proc Natl Acad Sci.*, **101:** 6309–6314 (2004).
- 54. Nagamine, T., Genic control of tolerance to chilling injury at seedling stage in rice. *Japanese Journal of Breeding*, **41:** 35-40 (1991).
- 55. Naidu, B., Thusitha, G. and Shu, F., Increasing cold tolerance in rice by selecting for high polyamine gibberellic acid content - A report for the Industries Research Rural Development Corporation (Australlian Government), RIRDC, Level 1, AMA House, 42-Macguarie Street BARTON ACT 2600,.BO Box 4776, KINGSTON ACT 2604 (2005).
- 56. Nakagahra, M., Okuno, K. and Vaughan, D., Rice genetic resources: History, conservation, investigative characterization and use in Japan. *Plant Molecular Biology*, **35:** 69-77 (1997).
- 57. Nishiyama, I., Effect of temperature on the vegetative growth of rice plants. In: Proceedings of the Symposium on Climate and Rice. IRRI pp. 159–186 (1976).
- 58. Nishiyama, I., Climatic influence on pollen formation and fertilization. *Biology of Rice*, **153**: 171 (1984).
- Nishiyama, I., Physiology of Cool Weather Damage to the Rice Plant. Sapporoo, Japan, Hokkaido University Press (1985).
- 60. Oidaira, H., Satoshi, S., Tomokazu, K. and Takashi, U., Enhancement of antioxidant enzyme activities in chilled rice seedlings. *Plant Physiology*, **156**: 811-813 (2000).
- 61. Pan, Y., Zhang. H., Zhang, D., Li, J., Xiong, H. and Yu, J., Genetic analysis of

- cold tolerance at the germination and booting stages in rice by association mapping. *PLoS ONE*, **10**: e0120590 (2015).
- 62. Park, I.K., Oh, C.S., Kim, D.M., Yeo, S.M. and Ahn, S.N., QTL Mapping for cold tolerance at the seedling stage using introgression lines derived from an intersubspecific cross in rice. *Plant Breeding and Biotechnology*, **1:** 1-8 (2013).
- 63. Peyman, S. and Hashem, A., Evaluation eighteen rice genotypes in cold tolerant at germination stage. *World Appl. Sci. J.*, **11:** 1476–1480 (2010).
- 64. Priyanka, K., Jaiswal, H.K., Waza, S.A. and Sravan, T., Response of rice seedlings to cold tolerance under boro conditions. *SABRAO Journal of Breeding and Genetics*, **47**: 185-190 (2015).
- 65. Ray, B.P., Rahman K.M., Hossain, M.E. and Sarker, P.C., DNA extraction protocol without liquid nitrogen of cold tolerant rice. *World Journal of Biology and Medical Sciences*, **3:** 59-67 (2016).
- 66. Sanghera, G.S., Wani, S.H., Hussain, W. and Singh, N.B., Engineering cold stress tolerance in crop plants. *Current Genomics*, **12:** 30-43 (2011).
- 67. Satya, P. and Saha, A., Screening for low temperature stress tolerance in boro rice. *IRRN*, **35**: (2010).
- 68. Shah, F., Huang, J., Cui, K., Nie, L., Shah, T., Chen, C. and Wang, K., Impact of hightemperature stress on rice plant and its traits related to tolerance. *Journal of Agricul-tural Science*, **149**: 545-556 (2011).
- 69. Sharifi, P., Evaluation on sixty-eight rice germplasms in cold tolerance at germination stage. *Rice Sci.*, **17**(1): 77–81 doi: 10.1016/S1672-6308(08)60107-9 (2010).
- Sheng, T., Dali, Z., Qian, Q. Yasufumi, K., Danian, H. and Lihuang, Z., QTL analysis of rice low temperature germinability. *Chinese Science Bulletin*, 46: 1800-1803 (2001).

- 71. Shimono, H., Hasegawa, T. and Iwama, K., Response of growth and grain yield in paddy rice to cool water at different growth stages. *Field Crops Research*, **73**: 67-79 (2002).
- 72. Shimono, H., Hasegawa, T., Fujimura, S. and Iwama. K., Responses of leaf photosynthesis and plant water status in rice to low water temperature at different growth stages. *Field Crops Research*, **89**: 71-83 (2004).
- 73. Song, S.Y., Chen, Y., Chen, J., Dai, X.Y. and Zhang, W.H., Physiological mechanisms underlying OsNAC5-dependent tolerance of rice plants to abiotic stress. *Planta*, **234**: 331–345 (2011).
- 74. Sonoike, K., The different roles of chilling temperatures in the photoinhibition of PSI and PSII. *J.Photochem. Photobiol. B. Biol.*, **48:** 136-141 (1999).
- 75. Sridevi, V. and Chellamuthu, V., Impact of weather on rice A review, International *Journal of Applied Research*, **1(9):** 825-831 (2015).
- 76. Sthapit, B.R. and Witcombe, J.R., Inheritance of tolerance to chilling stress in rice during germination and plumule greening. *Crop Science*, **38:** 660-665 (1998).
- 77. Su, C.F., Wang, Y.C., Hsieh, T.H., Lu, C.A., Tseng, T.H. and Yu, S.M., A novel MYBS3-dependent pathway confers cold tolerance in rice. *Plant Physiol.*, **153**: 145–158 (2010).
- 78. Thomashow, M.F., Plant cold acclimation; Freezing tolerance genes and regulatory mechanisms. *Annual Review of Plant Physiology & Molecular Biology*, **50:** 571-599 (1999).
- 79. Tian, Y., Zhang, H., Pan, X., Chen, X., Zhang, Z. and Lu, X. et al., Overexpression of ethylene response factor TERF2 confers cold tolerance in rice seedlings. *Transgenic Res.*, **20:** 857–866 (2011).
- 80. Ueno, K. and Miyoshi, M., Difference of optimum germination temperature of seeds of intact and dehusked japonica rice

- during seed development. *Euphytica*, **143**: 271–275 (2005).
- 81. Verheul, M.J., Picatto, C. and Stamp, P., Growth and development of maize (Zea mays L.) seedlings under chilling conditions in the field. *European Journal of Agronomy*, **5:** 31-43 (1996).
- 82. Watanabe, T, and Kume, T., A general adaptation strategy for climate change impacts on paddy cultivation: special reference to the Japanese context. *Paddy Water Environment*, **7:** 313-320 (2009).
- 83. Wise, R.R. and Naylor, A.W., Chilling induced photooxidation. Evidence for the role of singlet oxygen and superoxide in the breakdown of pigments and endogenous antioxidants. *Plant Physiology*, **83:** 278-282 (1987a).
- 84. Xiao, N., Huang, W.N., Zhang, X.X., Gao, Y., Li, A.L., Dai, Y., Yu. L., Liu, G.Q., Pan, C.H., Li, Y.H., Dai, Z.Y. and Chen, J.M., Fine mapping of qRC10-2, a quantitative trait locus for cold tolerance of rice roots at seedling and mature stages. *PLOS ONE*, **9:** e96046 (2014).
- 85. Xie, G., Kato, H. and Imai, R., Biochemical identification of the OsMKK6-OsMPK3 signaling pathway for chilling stress tolerance in rice. *Biochem. J.*, **443**: 95–102 (2012).
- 86. Xie, L., Zhengwei, T., Yuan, Z., Rongbao, X., Laibao, F., Yongzhong, X. and Xiaoquan, Q., Identification and fine mapping of quantitative trait loci for seed vigor in germination and seedling establishment in rice. *Journal of Integrative Plant Biology*, **56:** 749-759 (2014).
- 87. Xiong, L. and Yang, Y., Disease resistance and abiotic stress tolerance in rice are inversely modulated by an abscisic acidinducible mitogen-activated protein kinase. *Plant Cell*, **15**: 745–759 (2003).
- 88. Xu, M., Li, L., Fan, Y., Wan, J. and Wang, L., ZmCBF3 overexpression improves tolerance to abiotic stress in transgenic rice (Oryza sativa) without yield penalty. *Plant Cell Rep.*, **30:** 1949–1957 (2011).

- 89. Yang, A., Dai, X. and Zhang, W.H., A R2R3-type MYB gene, OsMYB2, is involved in salt, cold, and dehydration tolerance in rice. *J. Exp. Bot.* **63:** 2541–2556 (2012).
- 90. Yang, Z., Huang, D., Tang, W., Zheng, Y., Liang, K. and Cutler, A.J., Mapping of quantitative trait loci underlying cold tolerance in rice seedlings via high-throughput sequencing of pooled extremes. *PLOS ONE*, **8:** e68433 (2013).
- 91. Yoshida, R., Kanno. A., Sato, T. and Kameya, T., Cool temperature-induced chlorosis in. Rice plants. *Plant Physiology*, **110:** 997-1005 (1996).
- 92. Yoshida, S., Fundamentals of Rice Crop Science. Los Baños, Philippines, International Rice Research Institute (IRRI) (1978).

- 93. Yoshida, S., In: Fundamental of Rice Crop Science. IRRI, Los Banos pp.1–63 (1981a).
- 94. Yoshida, S., Climate environment and its influence on the rice plant. In:Fundamentals of Rice Crop Science. IRRI, Los Bonos pp. 65–110 (1981b).
- 95. Zhang, J., Li, J., Wang, X. and Chen, J., OVP1, a vacuolar H+-translocating inorganic pyrophosphatase (V-PPase), overexpression improved rice cold tolerance. *Plant Physiol. Biochem.* **49:** 33–38 (2011).
- 96. Zhang, Z.H., Su, L., Li, W., Chen, W. and Zhu, Y.G., A major QTL conferring cold tolerance at the early seedling stage using recombinant inbred lines of rice (Oryza sativa L.). *Plant Science*, **168**: 527–534 (2005).